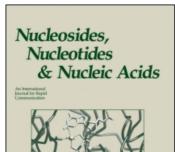
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SYNTHESIS AND ANTI-HIV EVALUATION OF 7-DEAZA ANALOGUES OF CARBOVIR

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Abstract: Three analogues of Carbovir 1 have been synthesized and evaluated for antiviral activity *in vitro*. Anti-HIV-1 and anti-HIV-2 activities have been observed with 7-deaza analogues 3 and 5 of 1. Compound 5 was about ten times more potent than 3 against HIV-1 and HIV-2 on different cell lines.

Introduction

Carbovir (1) has emerged as the first carbocyclic nucleoside analogue with potential as a therapeutic agent for the treatment of AIDS¹. Furthermore some 2,3-dideoxy-2,3-didehydro-pyrrolo[2,3-d] pyrimidine nucleosides in their triphosphate form such as 2² have been shown to be potent and specific inhibitors of the reverse transcriptase (RT) of HIV-1.

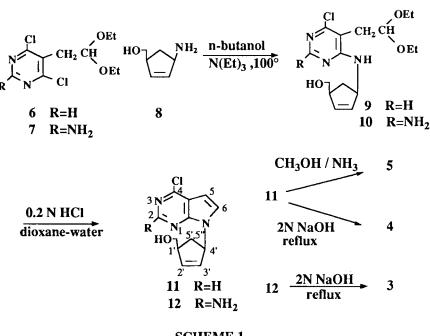
Therefore, in an attempt to increase selectivity and potency against HIV RT, we have been interested to combine their structural features in the same molecule.

Chemistry

We have synthesized three new analogues of Carbovir as racemic mixtures, in which the 7-nitrogen atom of the purine ring has been deleted, providing 7-deaza guanine (3), 7-deazahypoxanthine (4) and 7-deazaadenine (5) analogues of Carbovir (1).

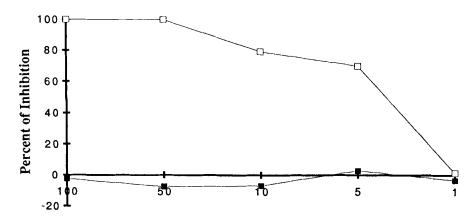
$$H_{2}N$$
 $H_{2}N$
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 H_{3}
 H_{4}
 H_{5}
 H

Pyrimidines 6³ and 7⁴ were reacted with cyclopentenylamine 8⁵ in butanol to provide the carbocyclic pyrimidines 9 and 10 respectively (Scheme 1). Cyclisation was then performed in dilute hydrochloric acid at room temperature to give 11 and 12. Nucleophilic displacement of the chlorine atom in 11 and 12 was then carried out with sodium hydroxide to give 4 and 3, whereas treatment of 11 with liquid ammonia gave 5.



SCHEME 1

Inhibition of HIV-1 RT in CEM-SS Cell Culture Supernatant



Concentration of Compound 5 (µM)

FIG.1.The percentage of RT inhibition is expressed in comparison with untreated cells which were incubated in the presence the solvent (DMSO); ---: RT inhibition; ---: cytoxicity.

Biology

Compounds 3, 4, 5, 11 and 12 were tested for anti-HIV-1 activity on CEM-SS cells. Compound 5 exhibited a significant inhibitory property and no toxicity as shown on Fig. 1 while compound 3 also inhibited HIV replication but was less active.

These two compounds were further analyzed for their activity on different isolates of HIV-1 and HIV-2 and on different cells including primary cultures of PBMC. Non. of these molecules were toxic on these cells at the highest concentration used (10^{-4} M). Table 1 indicates the IC₅₀ measured for HIV-1 and HIV-2. Compounds 5 and 3 were 10 times more active on PBMC than on cell lines and they inhibited HIV-1 and HIV-2 with the same efficiency. Compound 5 with an IC₅₀ of 3 10^{-6} M on CEM-SS cells is slightly less potent than Carbovir 1 (IC₅₀ of 6 10^{-7} M)¹.

Compounds 3, 4, 5, 11 and 12 were found inactive against HSV-1 HSV-2, HCMV, Vaccinia virus, VSV, and Sindbis virus. Compound 3 has been reported recently by W. Schneller et al⁶ and was also found inactive against HSV-1, HSV-2, HCMV, and HIV-1 in CEM cells, different from the CEM-SS used in this work.

In conclusion, this series of compounds is non-cytotoxic and two of them, the 7-deaza adenine (5) and the 7-deaza guanine (3) analogues were found active against HIV-1 and HIV-2 in vitro, although slightly less active than Carbovir.

TABLE 1

ANTI-HIV ACTIVITY
INHIBITORY CONCENTRATION 50 (IC50) OF ACTIVE MOLECULES

VIRUS/CELLS	3		5	
HIV-1 LAI/CEM-SS	1.5 10	-5 M	3	10 ⁻⁶ M
HIV-1 LAI/CEMX174	2.5 10	-5 M	2.5	10 ⁻⁶ M
HIV-1 IIIB/PBMC	7 10	-6 M	4	10 ⁻⁷ M
HIV-2 D194/PBMC	6 10	-6 M	3.5	10 ⁻⁷ M
HIV-2 D205/PBMC	9 10	-6 M	3.5	10 ⁻⁷ M

Experimental section

Chemistry

The melting points were taken on a Kofler hot stage apparatus and were uncorrected. Elemental analyses were performed by the "Service de Microanalyse", CNRS, ICSN, 91198 Gif-sur-Yvette, France. Fast atom bombardment (FAB) mass spectra (MS) were obtained from the "Laboratoire de Spectrometrie de Masse", CNRS, ICSN, 91198 Gif-sur-Yvette, France. Proton nuclear magnetic resonance (1H-NMR) spectra were recorded at 200 MHz on a Bruker AC 200 spectrometer. Numbering of cyclopentene methanol in compounds 5, 9, 10, 11, and 12 and of the pyrrolo[2,3-d]pyrimidine ring is indicated for 11 and 12 on scheme 1 as well as the numbering of cyclopentenyl derivatives of pyrrolo[2,3-d]pyrimidinones 3 and 4 which is mentioned for 3.

(\pm)-[cis-4 -(4-Chloro-7H-pyrrolo[2,3-d]pyrimidin-7-yl)-cyclopent-2 -enyl]-methanol (11).

A solution of 6 (9.7 g, 36.6 mmol) in n-butanol (50 ml) was treated with 8⁵ (3.2 g, 28.31 mmol) in the presence of an excess of triethylamine (4 ml) at 100°C for 3 days under nitrogen. The mixture was then cooled, evaporated to dryness and coevaporated several times in the presence of n-heptane. The residue was partitioned between chloroform and water. The organic layer was washed twice with water, dried (MgSO₄), filtered and concentrated under reduced pressure. The crude product (11.34 g) was purified by column chromatography (silica gel, CH₂Cl₂-EtOH 19:1) to give 5.4 g of

unreacted pyrimidine 6 and 5.5 g of 9 as an oil which was contaminated with about 10 % of 11 according to its 1 H NMR spectrum in CDCl₃. 9 : 1 H NMR (CDCl₃) δ 8.27 (s, 1H, H-2) ; 6.41 (d, 1H, NH, $J_{NH-4'}$ = 7.4 Hz) ; 5.87 (m, 2H, \underline{CH} = \underline{CH}) ; 5.19 (m, 1H, H-4') ; 4.59 (t, 1H, \underline{CH} (OEt)₂, JCH-CH₂-pyrimidine = 5.4 Hz) ; 3.57 (m, 6H, 2 x O<u>CH</u>₂CH₃, \underline{CH}_{2} OH) ; 2.91 (m, 3H, \underline{CH}_{2} -pyrimidine, H-1') ; 2.65 (m, 1H, H5') ; 1.46 (m, 1H, H5") ; 1.21 (m, 6H, 2 x OCH₂CH₃). FAB-MS m/e = 342

A solution of **9** obtained as described above (2.74 g, 10.33 mmol) in dioxane-1N aqueous hydrochloric acid (4 : 1) was stirred at room temperature for 2 days. An excess of concentrated ammonium hydroxide was then added and the mixture was evaporated under reduced pressure. The residue was partitioned between dichloromethane and water. The organic layer was washed twice with water, dried (MgSO₄), filtered and evaporated to dryness. The oily residue crystallized and was recrystallized from a mixture of cyclohexane-ethyl acetate (4 : 1) to give a 87 % yield of 11; mp 95-96°C; 1 H NMR (DMSO-d₆) δ 8.66 (s, 1H, H2); 7.69 (d, 1H, H6, J $_{6-5}$ = 3.65 Hz); 6.69 (d, 1H, H5, J $_{5-6}$ = 3.65 Hz); 6.19 (m, 1H, H2', J $_{3'-2'}$ = 5.5 Hz, J $_{2'-4'}$ = 2 Hz, J $_{2'-1'}$ = 2 Hz); 5.95 (m, 1H, H4', J $_{4'-3'}$ = 2.1 Hz, J $_{4'-5'}$ = 5.85 Hz, J $_{4'-5''}$ = 8.7Hz); 5.87 (m, 1H, H3', J $_{3'-1'}$ = 2.1 Hz); 4.87 (t, 1H, OH, JOH-CH₂= 5.5 Hz); 3.5 (m, 2H, CH₂OH); 2.94 (m, 1H, H1', J $_{1'-5'}$ = 8.7 Hz, J $_{1'-5''}$ = 5.85 Hz); 2.71 (m, 1H, H5', J $_{5'-5''}$ =13.55 Hz); 1.60 (m, 1H, H5''). Anal. Calcd for C₁₂H₁₂N₃O Cl: C, 57.72; H, 4.84; N, 16.83; Cl, 14.20. Found: C, 57.69; H, 4.66; N, 16.74; Cl, 14.43.

(\pm)-cis-7-[4 -(Hydroxymethyl)-cyclopent-2 -enyl]-3H,7H-pyrrolo[2,3-d] pyrimidin-4-one (4).

A solution of **11** (327 mg, 1.3 mmol) in 2N NaOH (20 ml) was refluxed for 5 hours. The mixture was neutralized with acetic acid, evaporated to dryness and coevaporated several times with toluene. The residue was adsorbed onto silica gel and purified by column chromatography eluting with dichloromethane-ethanol (19 : 1). Yield 220 mg (73 %); an analytical sample was obtained by crystallization from water. mp. 86-87°C; ¹H NMR (DMSO-d₆) δ 11.90 (s, 1H, NH); 7.93 (s, 1H, H2); 7.06 (d, 1H, H6, J₆₋₅ = 3.5 Hz); 6.52 (d, 1H, H5, J₅₋₆ = 3.5 Hz); 6.14 (m, 1H, H3', J_{2'-3'} = 5.5 Hz, J_{3'-1'} = 2 Hz, J_{3'-4'} = 2Hz); 5.84 (m, 1H, H2', J_{2'-1'} = 2.1 Hz; J_{2'-4'} = 2.1 Hz); 5.77 (m, 1H, H1', J_{1'-5'} = 8.6Hz, J_{1'-5''} = 6 Hz); 4.75 (t, 1H, OH, J = 5 Hz); 3.48 (t, 2H, <u>CH₂OH</u>, JCH₂OH = 5 Hz, JCH₂-4'= 6 Hz); 2.90 (m, 1H, H4', J_{4'-5'} = 8.6 Hz, J_{4'-5''} = 6 Hz); 2.65 (m, 1H, H5', J_{5'-5''} = 13.6 Hz); 1.54 (m, 1H, H5''). Anal. Calcd for C₁₂H₁₃N₃O₂: C, 62.33; H, 5.67; N, 18.17. Found: C, 62.01; H, 5.52; N, 18.01.

(±)-[cis-4 -(4-Amino-7H-pyrrolo[2,3-d]pyrimidin-7-yl)-cyclopent-2 -enyl]-methanol (5).

A solution of **11** (700 mg, 2.8 mmol) in methanol (70ml) was treated with liquid ammonia (70 ml) at 100°C in a stainless sealed vessel for 48 hours. The mixture was evaporated to dryness, adsorbed onto silica gel, and purified by silica gel column chromatography using dichloromethane-ethanol (19:1) as eluent affording 640 mg of a colorless oil (yield, 99 %). Analytically pure material was obtained by crystallization from water. mp 170-172°C; 1 H NMR (CDCl₃) δ 8.28 (s, 1H, H₂); 7.04 (d, 1H, H₆, J₆₋₅ = 3.6 H₂); 6.31 (d, 1H, H₅); 6.09 (m, 1H, H₂', J_{2'-3'}= 5.5 H₂, J_{2'-1'} = 2.1 H₂); 5.83 (m, 1H, H₃', J_{3'-1'} = 2.18); 5.73 (m, 1H, H₄', J_{4'-3'}= 2.2 H₂; J_{4'-5}= 9.1 H₂; J_{4'-5'}= 6.7 H₂; J_{4'-2'}= 2.1 H₂); 5.08 (s, 2H, NH₂); 3.78 (m, 2H, CH₂OH, 2 J_{6'-6"}= 10.66 H₂, J_{6'-1'} = 4.25 H₂); 3.07 (m, 1H, H₁', J_{1'-2'} = 2.1 H₂, J_{1'-3'}= 2.18 H₂, J_{1'-5'}= 9.1 H₂, J_{1'-5'}= 6.7 H₂, J_{1'-6'}= J_{1'-6''}= 4.25 H₂); 2.79 (m, 1H, H₅', 2 J_{5'-5"}= 14 H₂); 1.95 (m, 1H, H₅''). Anal. Calcd. for C₁₂H₁₄N₄O: C, 62.59; H, 6.13; N, 24.33. Found: C, 62.73; H, 5.81; N, 23.97.

(±)-[cis-4 -[[2-Amino-6-chloro-5-(2,2-diethoxy-ethyl)-pyrimidin-4-yl]-amino]cyclopent-2 -enyl]-methanol (10).

A solution of **7** (2.77 g, 9.89 mmol) in n-butanol (30 ml) was treated with **8** (1.7 g, 15 mmol) according to the procedure used in the preparation of compound **9**. After extraction, a solid residue was obtained and purified by column chromatography (silica gel) eluting with dichloromethane-ethanol (49 : 1). Crystallization from ethanol afforded 1.4 g (40 %) of **10**; mp 153-154°C; ¹H NMR (CDCl₃) δ : 6.19 (d, 1H, NH, J_{NH-4}; = 7.54 Hz); 5.84 (m, 2H, CH = CH); 5.08 (m, 1H, H4'); 4.84 (s, 2H, NH₂); 4.48 (t, 1H, CH₂CH(OEt)₂; 3.75-3.40 (m, 6H, 2xOCH₂CH₃, CH₂OH); 2.90 (m, 1H, H1'); 2.7 (m, 2H, CH₂CH(OEt)₂; 2.54 (m, 1H, H5'); 1.42 (m, 1H, H5"). Anal. Calcd for C₁₆H₂₅N₄O₃Cl: C, 53.85; H, 7.06; N, 15.70. Found: C, 54.11; H, 6.94; N, 15.87. FAB-MS m/e = 357.

(\pm)-[cis-4 -[2-Amino-4-chloro-7*H*-pyrrolo[2,3-d]pyrimidin-7-yl]-cyclopent-2 -enyl]-methanol (12).

Compound 10 was treated according to the procedure used in the preparation of compound 11 to give 12 with a yield of 76%; mp (CHCl₃) 144-145°C; ¹H NMR (DMSO-d₆) δ : 7.10 (d, 1H, H₆, J₆₋₅ = 3.75 Hz); 6.67 (s, 2H, NH₂); 6.35 (d, 1H, H₅); 6.13 (m, 1H, H₂', J_{3'-2'} = 5.5 Hz; J_{2'-1'} = 2 Hz); 5.82 (m, 1H, H₃', J_{3'-4'} = 2 Hz); 5.69 (m,

1H, H4', $J_{4'-3'}=2$ Hz, $J_{4'-5'}=6$ Hz, $J_{4'-5''}=8.7$ Hz); 4.76 (t, 1H, OH, JOH-CH₂= 5.3 Hz); 3.49 (m, 2H, CH₂OH, $J_{1'-6''}=J_{1'-6''}=2$ Hz); 2.90 (m, 1H, H1', $J_{1'-5}=8.7$ Hz); 2.60 (m, 1H, H5'); 1.55 (m, 1H, H5'', $J_{5'-5''}=13.6$ Hz). Anal. Calcd. for $C_{12}H_{13}N_4OCl.: C$, 54.45; H, 4.95; N, 21.16. Found: C, 54.32; H, 4.95; N, 20.91.

(±)-2-Amino-7-[cis-4 -(hydroxymethyl)-cyclopent-2 -enyl]-3H,7H-pyrrolo[2,3-d]pyrimidin-4-one (3).

Compound 12 was treated according to the procedure used in the preparation of compound 4 to give 3 with a yield of 56%; The titled compound crystallized from waterethanol (4 : 1) after column chromatography (silica gel), eluting with dichloromethaneethanol (19 : 1). mp: decomposition started at 254-255°C and was complete at 270°C; 1 H NMR (DMSO-d₆) δ 10.30 (s, 1H, NH); 6.64 (d, 1H, H₆, J₆₋₅= 3.6 Hz); 6.27 (d, 1H, H₅); 6.22 (s, 2H, NH₂); 6.09 (m, 1H, H₃', J_{3'-2'}= 5.6 Hz, J_{3'-4'}= 2.06 Hz, J_{3'-1'} = 2.06 Hz); 5.77 (m, 1H, H2', J_{2'-1'}= 2.14 Hz, J_{2'-4'}= 2.14 Hz); 5.57 (m, 1H, H1', J_{1'-5'}= 8.7 Hz, J_{1'-5''}= 6.2 Hz); 4.74 (t, 1H, OH, JOH-CH₂= 5 Hz); 3.46 (t, 2H, CH₂OH, JCH₂-H_{4'}= 5.6 Hz); 2.86 (m, 1H, H4', J_{4'-5''}= 8.7 Hz, J_{4'-5''}= 6.2 Hz); 2.5 (m, 1H, H5'); 1.48 (m, 1H, H5'', J_{5'-5''}= 13.5 Hz). Anal. Calcd for C₁₂H₁₄N₄O₂: C, 58.53; H, 5.73; N, 22.75. Found: C, 58.64; H, 5.71; H, 22.44.

Biology

Cell cultures and viruses

The human T-cell lines CEM-SS⁸, CEM-X 174 fusion of a B-cell line (obtained through the AIDS Research and Reference Reagent Program-Division of AIDS, NIAID, NIH from Cresswell, P.) were maintained in RPMI 1640 medium with 2 mM glutamine and 10 % fœtal bovine serum (FBS, heated 30 min. at 56°C) in a CO₂ incubator at 37°C. Human peripheral blood mononuclear cells (PBMC) from HIV seronegative donors, separated on ficoll, were cultured in RPMI 1640 containing 10 % FBS in the presence of 4 μ l/ml phytohaemagglutinin (PHA). After 3 days, PHA was removed, cells were washed with medium and reincubated, after infection, in medium containing interleukin-2 (20 U/ml). Several isolates of HIV were used; HIV-1 LAI^{9,10}, HIV-1 IIIB¹¹, HIV-2 D194 and HIV-2 D205¹².

Inhibition of virus production

Production of virus particles was evaluated by measurement of virion associated reverse transcriptase activity in the culture supernatants as described (Jonassen, T.O., Skatron Application Notes, April 1986).

Cells were infected with the HIV isolates (20 TC:D₅₀). After 0.5 h adsorption, the residual free virus was removed by centrifugation, the cultures were resuspended at the

concentration of 40×10^3 cells/ml for CEM-SS, 80×10^3 for CEM -X 174, 160×10^3 in the medium indicated above and distributed into microtitration plates (100μ l/well) containing 0.1 ml/well of different dilutions of the antiviral drugs. Reverse transcriptase activity was measured after 5 days for CEM-SS and CEM-X 174, or after 7 days for PBMC. In this later case, 100μ l of medium was removed at day 5 and replaced with fresh medium containing the same concentration of drug. Fifty per cent inhibitory concentration (IC₅₀) was defined as the concentration of drug that reduced the reverse transcriptase activity by 50 %. Cytotoxicity was determined by the MTT dye reduction assay¹³.

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